

X-ray absorption spectroscopy of Zn sites in biology

When is a thiolate not a thiolate?

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Zn is one of the most widespread elements in biology

- Zn is typically coordinated to S (thiolate), N (Histidine) or O (water, carboxylate)
- Zn is usually tetrahedral
- Question of interest is determining the relative number of S vs. N/O ligands

EXAFS can readily distinguish S from N/O ligation

- EXAFS can distinguish ligands that differ by ca. 10 in atomic number

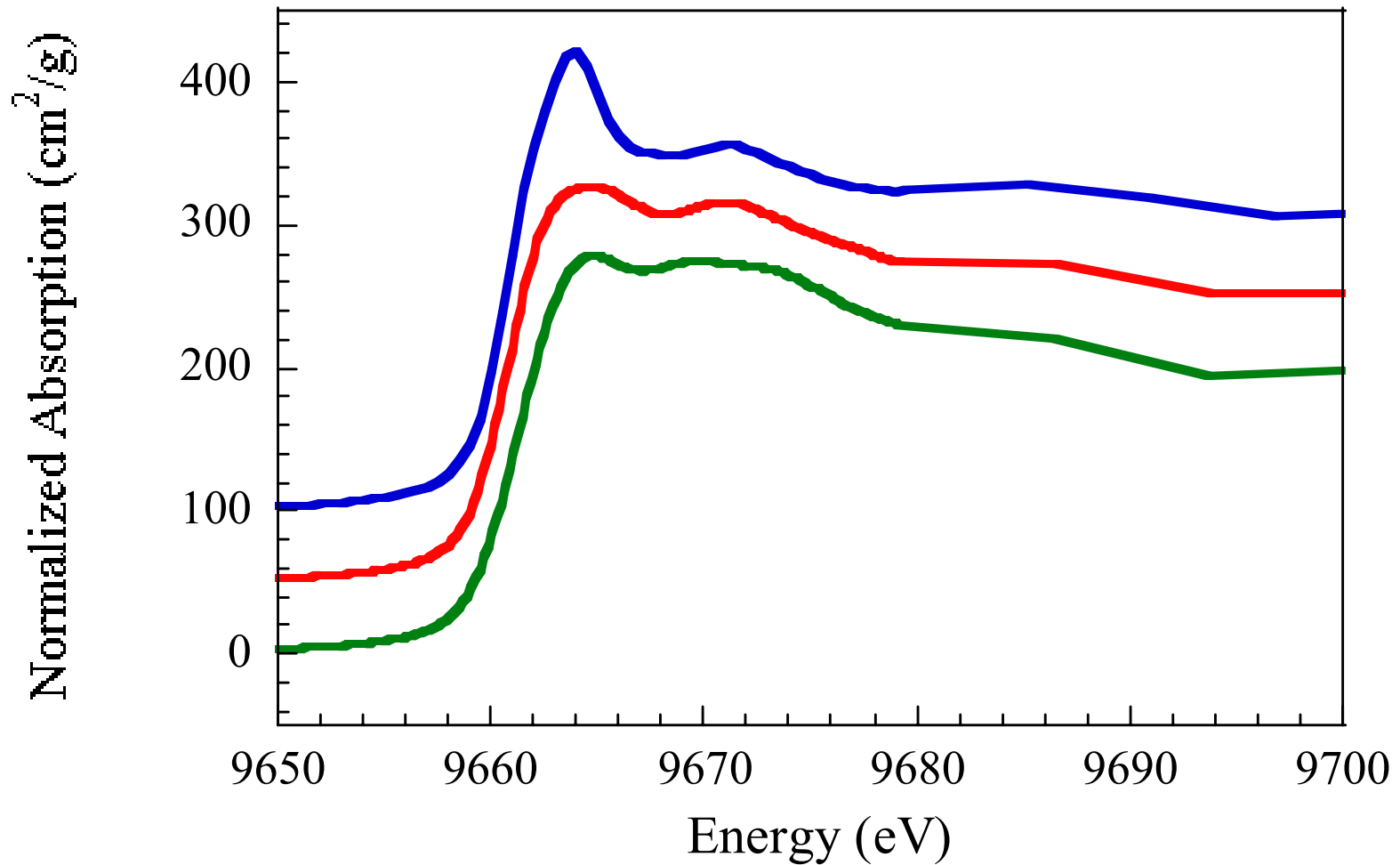
However

- There are several examples in which EXAFS has overestimated the number of sulfur ligands.
- And there are cases where EXAFS has underestimated the number of sulfur ligands.

Goal: Use well-defined model compounds to understand limitations of ligation determination

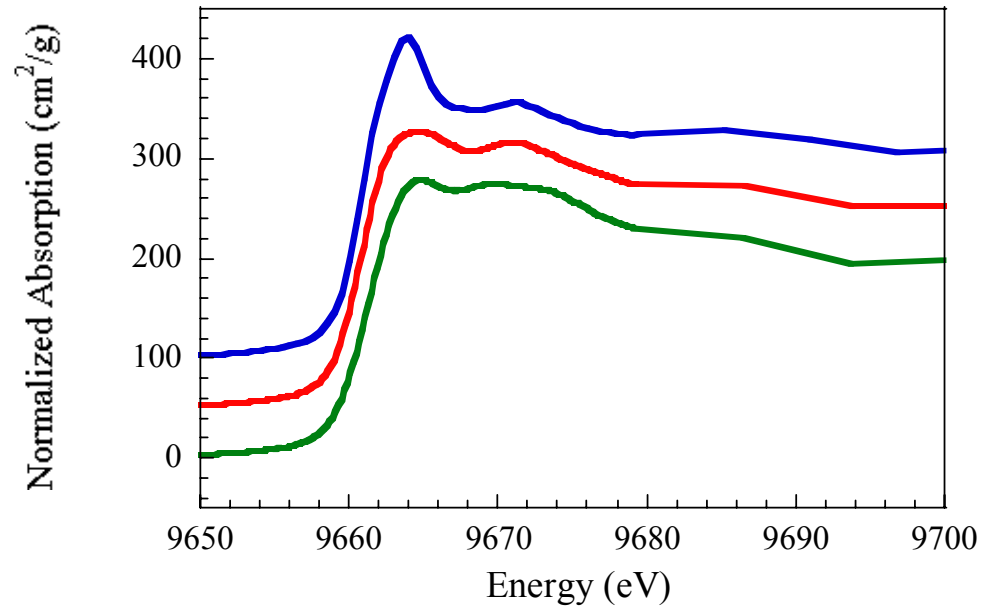
- “Inorganic” – thiolate/imidazole models (S. Koch).
- “Peptide” – cysteine/histidine peptide models (J. Berg)

XANES spectra depend on Zn ligation

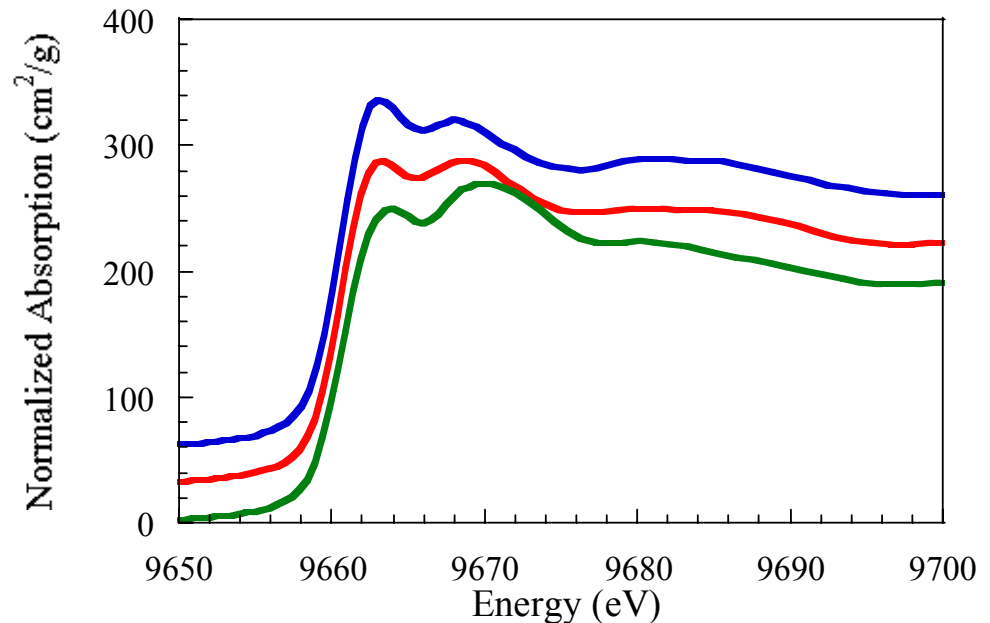


However, variation from sample to sample is larger than variation from ligation to ligation. (Despite the fact that 9660 eV feature is sometimes identified as “characteristic of ZnS₄ ligation.

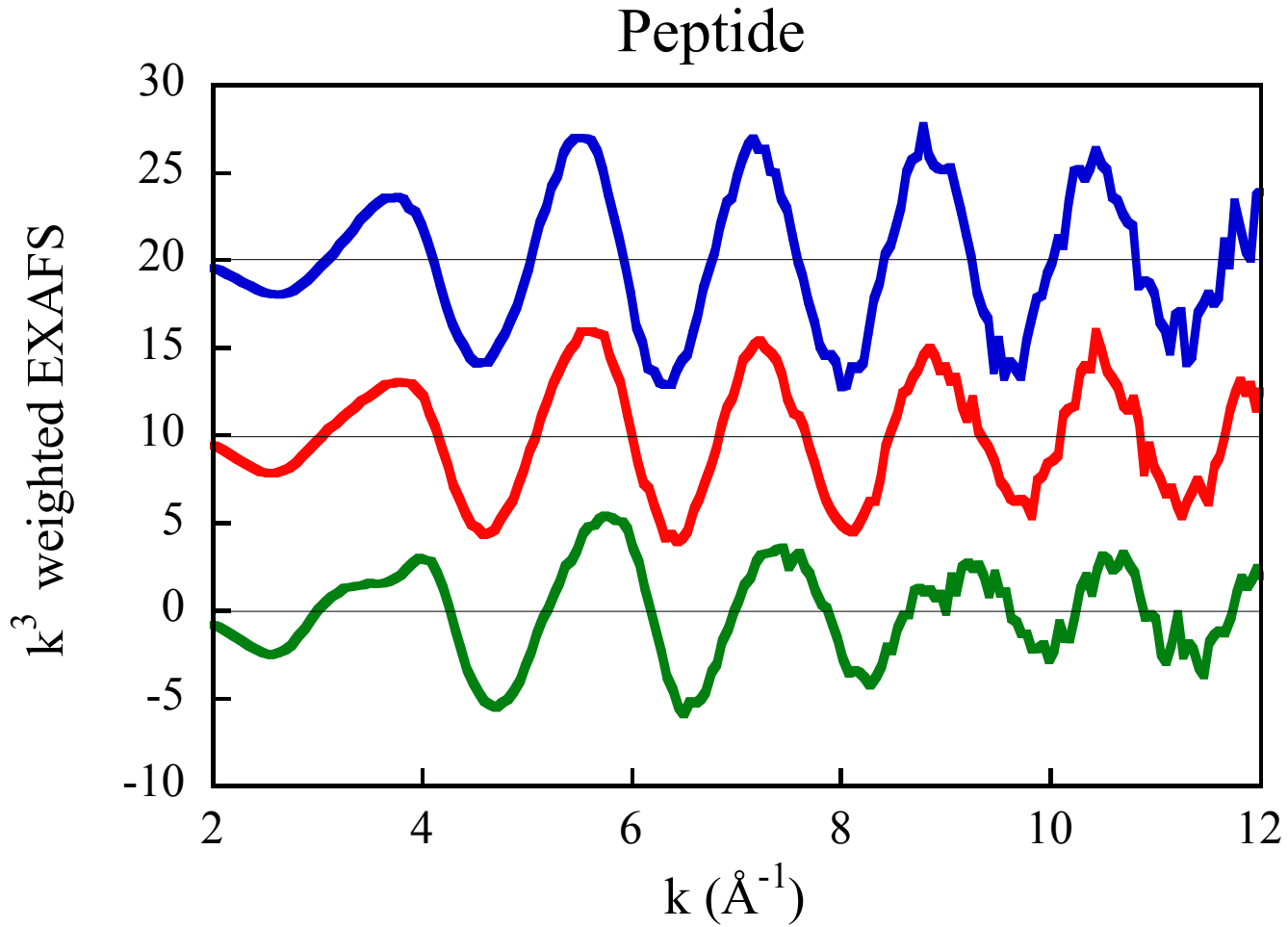
Inorganic edges



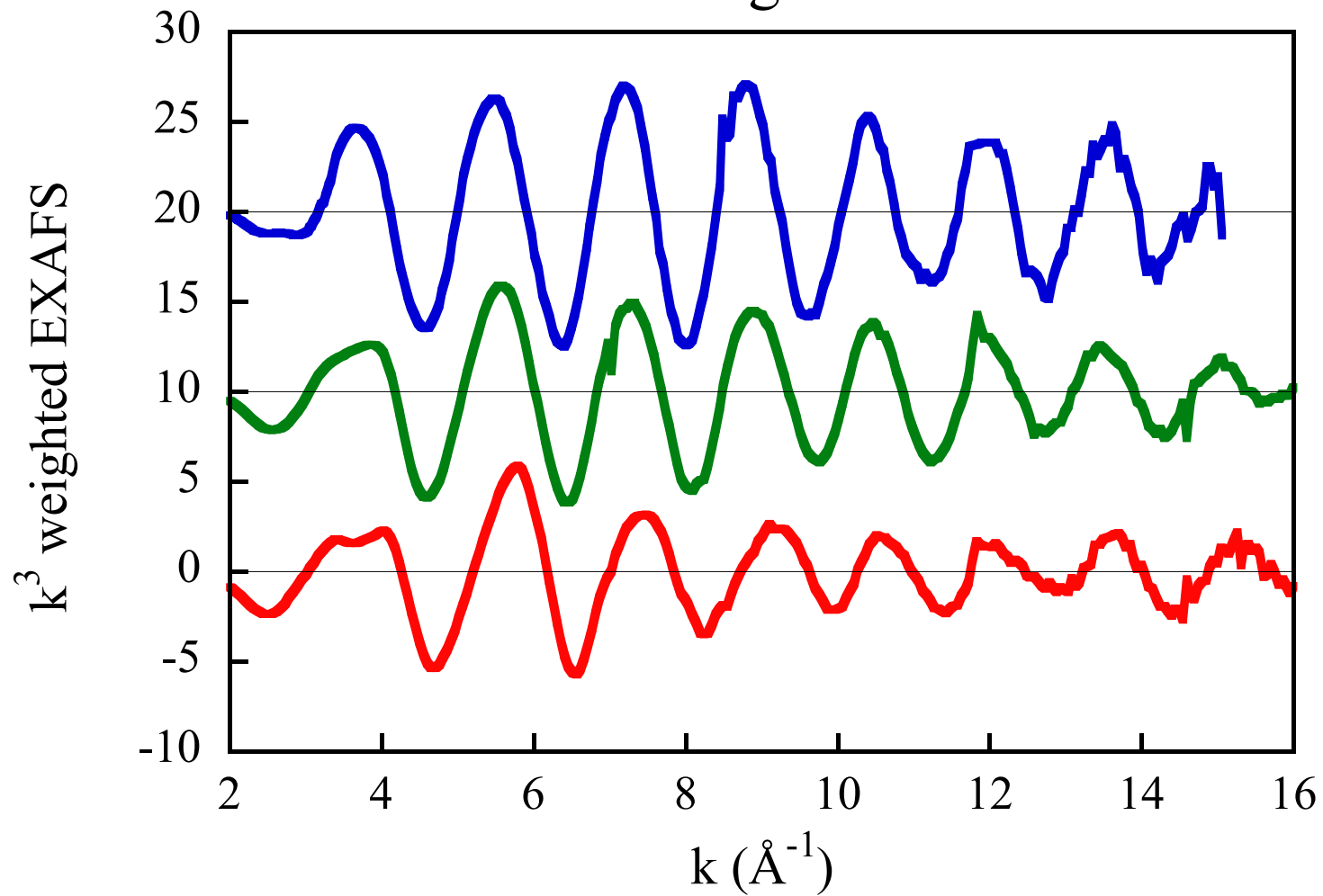
Peptide edges



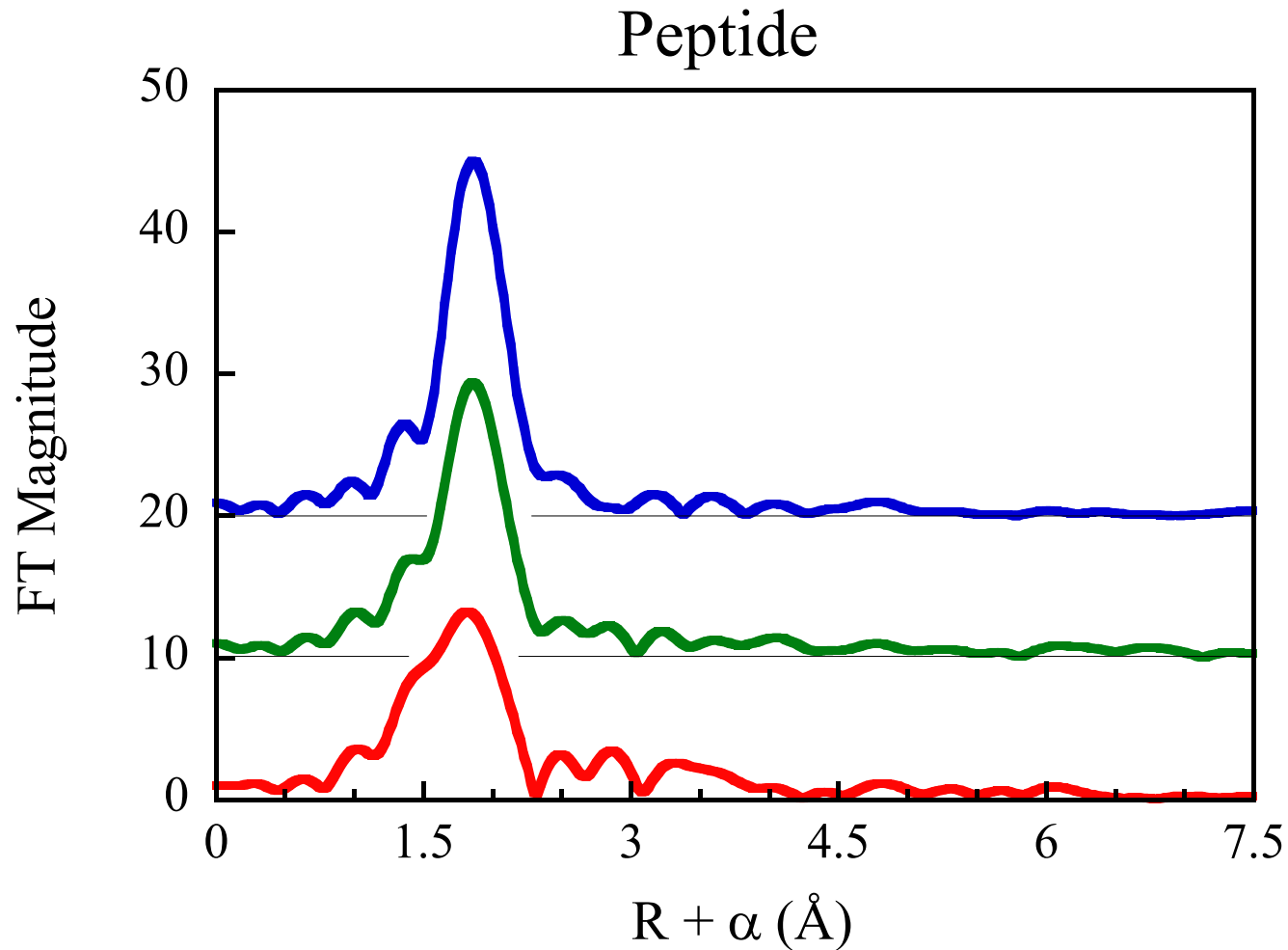
EXAFS shows only small amplitude changes



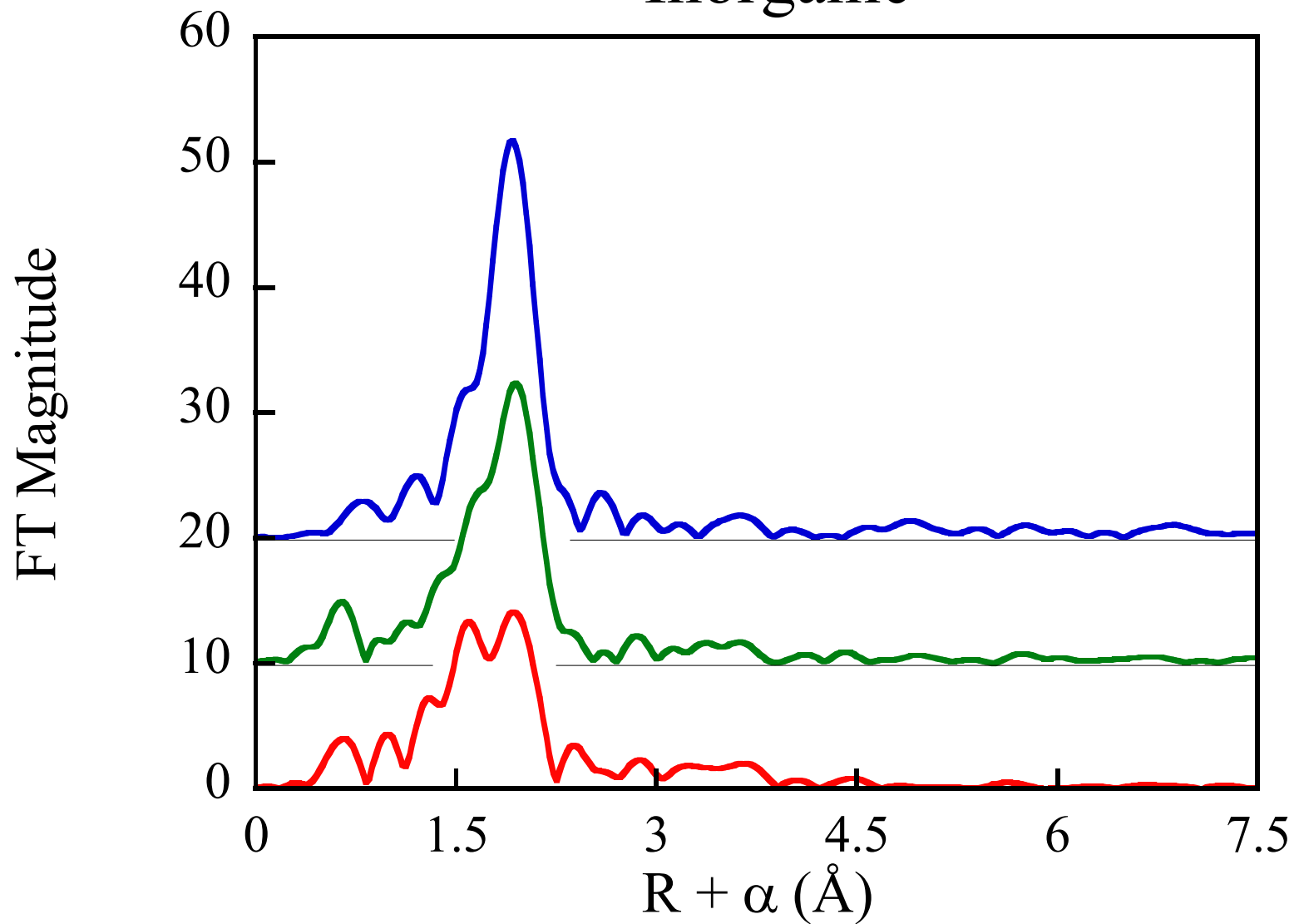
Inorganic



FTs do not show 2 obvious shells



Inorganic

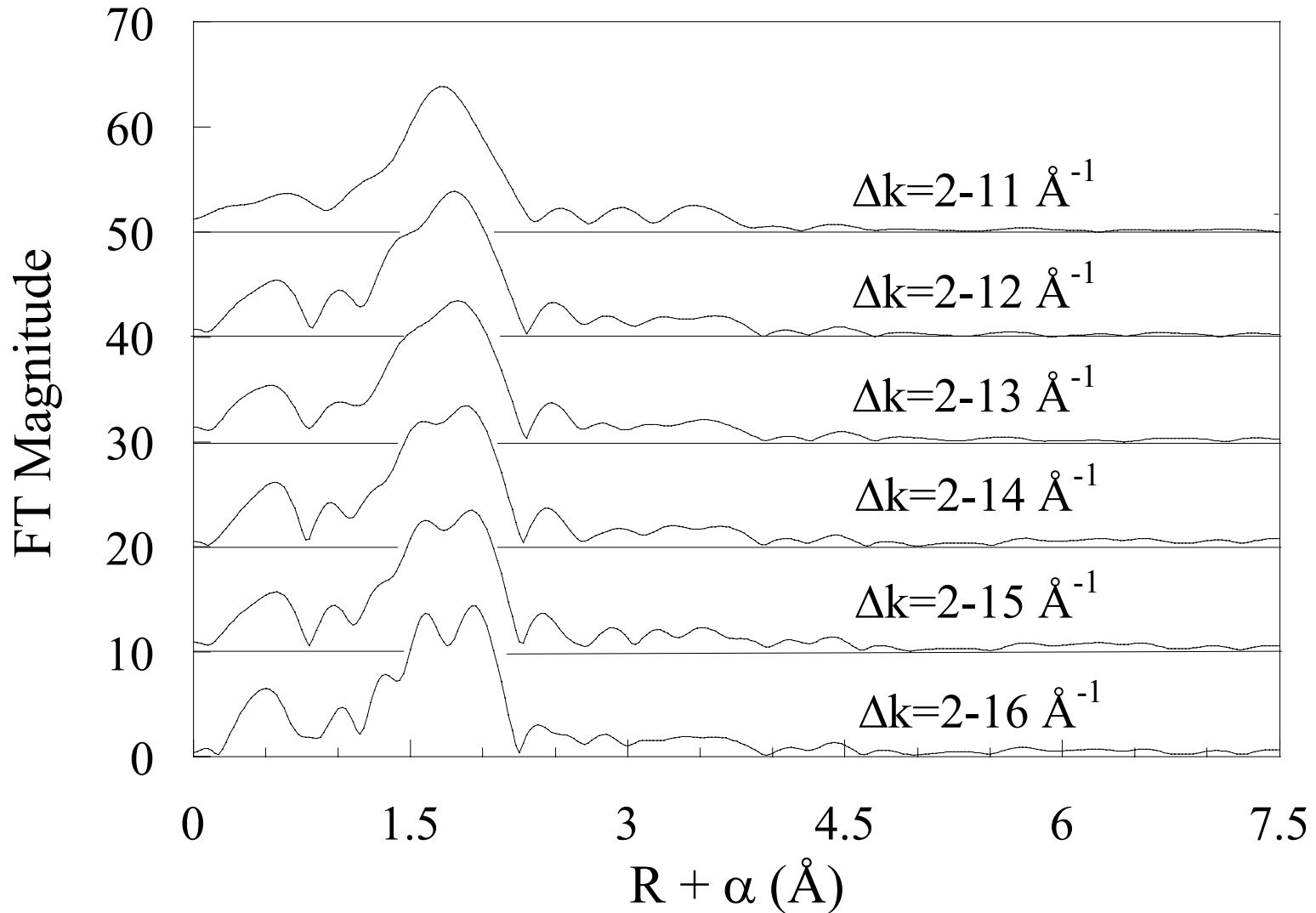


The lack of resolvable peaks is a consequence of:

- Unfortunate distances ($Z_n-N=2.05$; $Z_n-S=2.3$)
- Destructive interference -- $\varphi_N \approx \varphi_S + \pi$

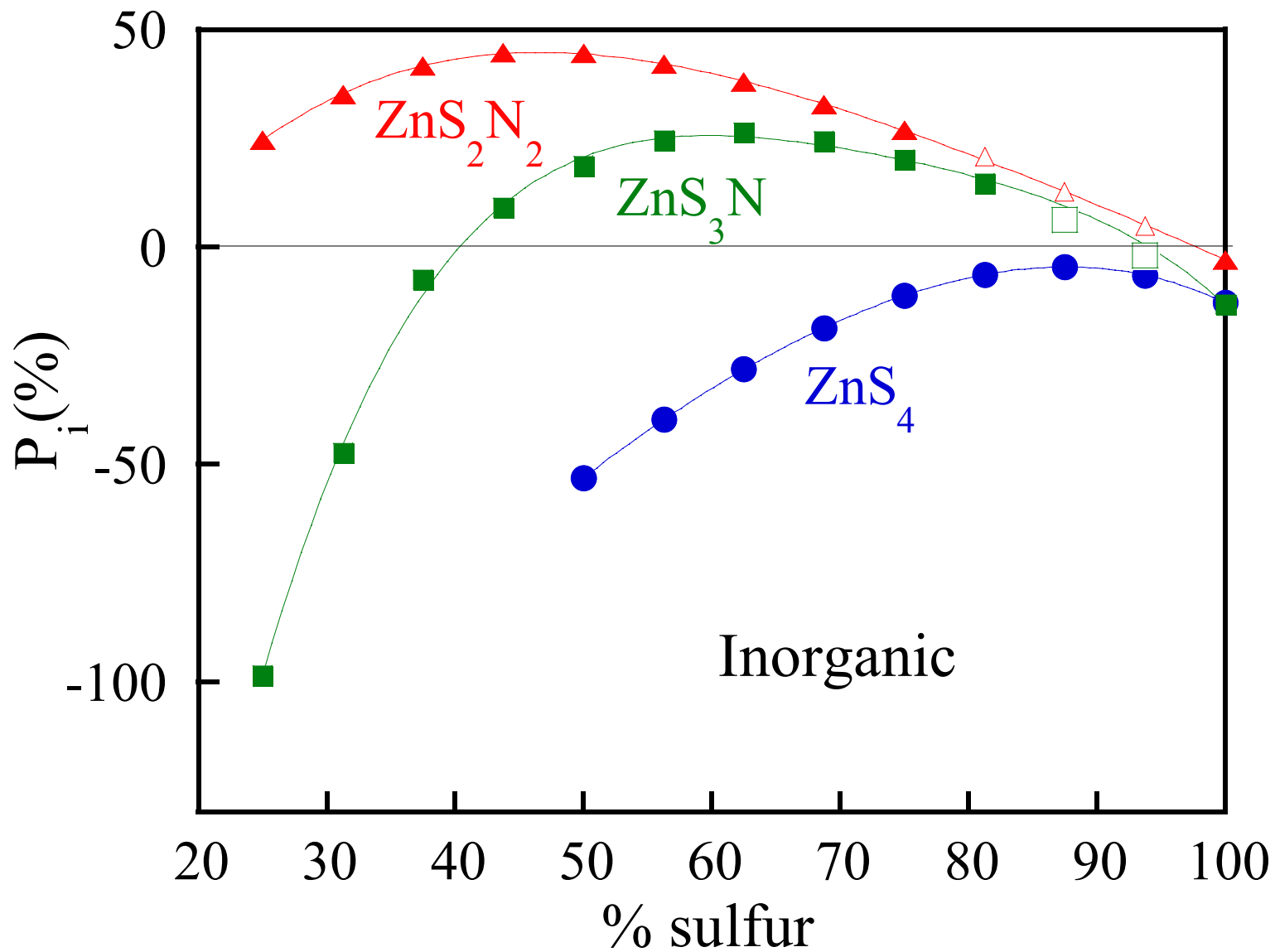
This has the result that S and N oscillations are nearly out of phase for much of the accessible k range

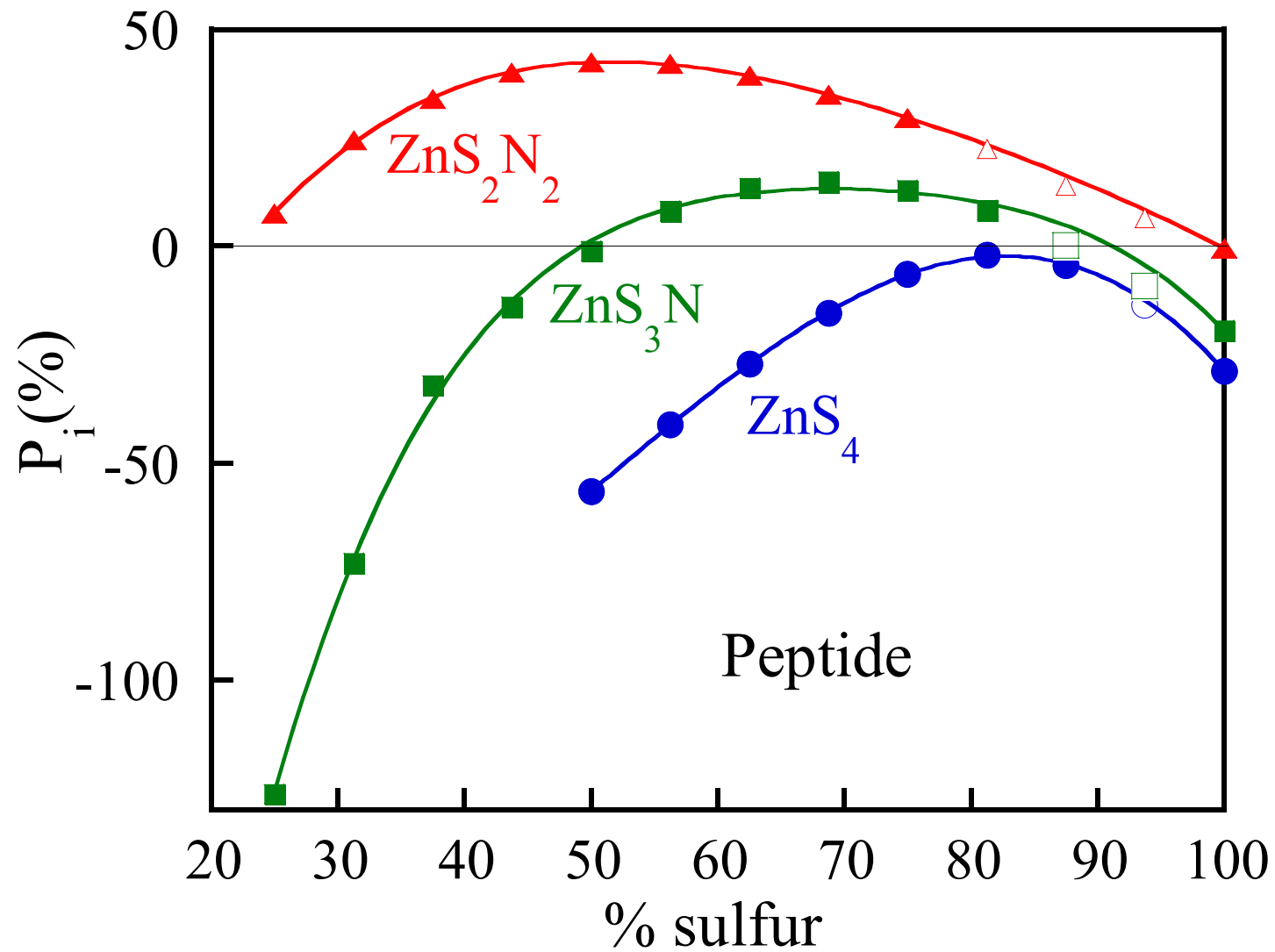
One solution is to measure data over wide k range
(ZnS_2N_2 inorganic)



Treat %S as a continuous variable

- Define P_i as percent improvement in fit.
- To avoid changes in degrees of freedom, P_i defined with respect to a fictitious ZnS_2S_2 fit.
- Dependence of P_i on %S is highly characteristic of ligation – $P_i=0$ means that mixed ligation does not give any improvement in the fit.

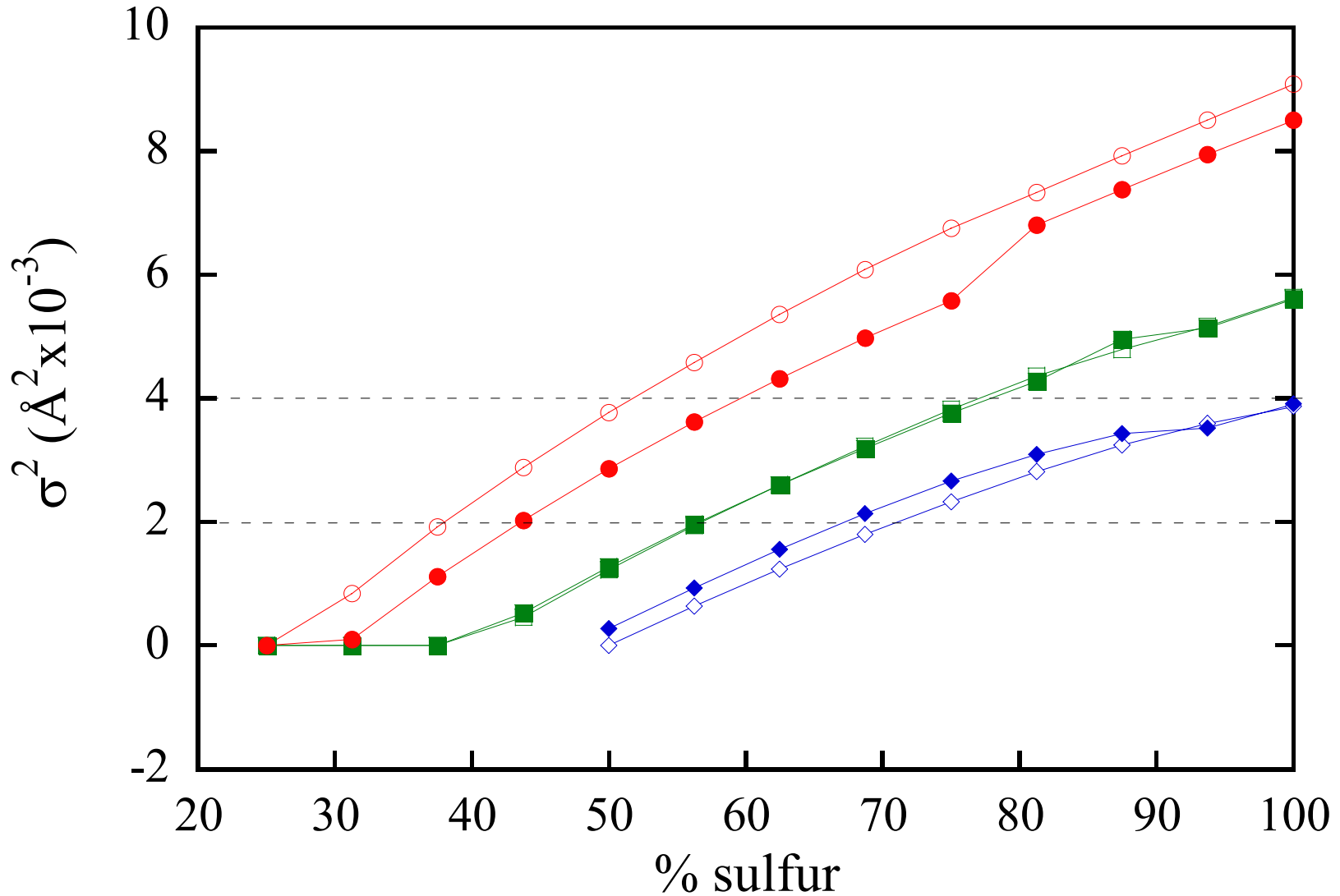




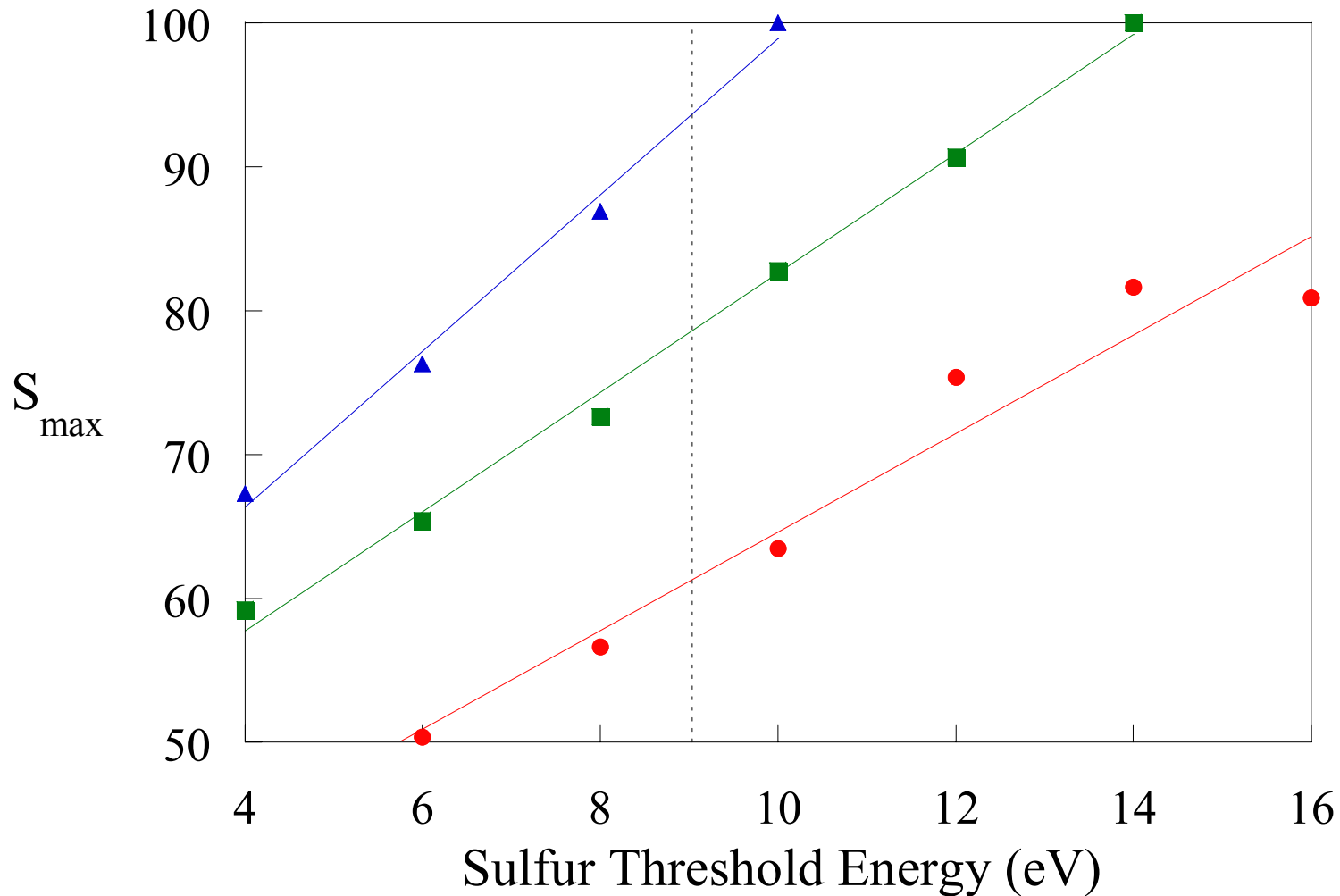
Dependence of P_i on %S

- Maximum in P_i approximately matches expected %S.
- The height of the maximum in P_i increases for samples that have authentic mixed ligation.
- P_i always increases when a nitrogen component is added.

In addition to P_i , σ^2 depends on ligation



S_{\max} (% S giving optimum P_i) depends on E_0



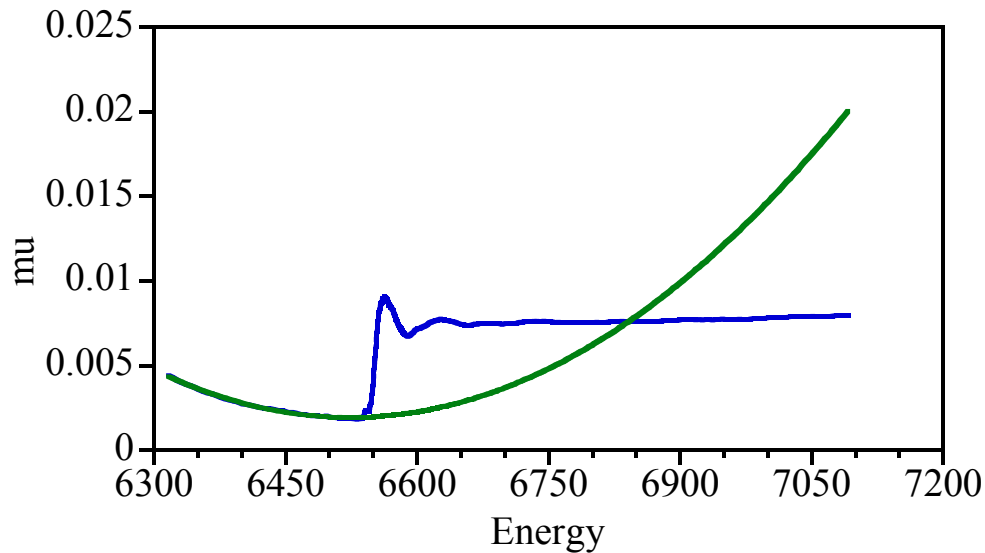
E_0 can have pronounced effect on apparent ligation

- Sensitivity to E_0 is a consequence of the fact that the difference between S and N is largely encoded in their phase difference.
- E_0 is often been treated as a freely variable parameter.
- Range of “chemically reasonable” variation has been given as ± 20 eV.
- Variation by of E_0 by more 3-4 eV from calibrated value (9 eV) changes the apparent ligation.

XANES spectra contain useful information regarding structure

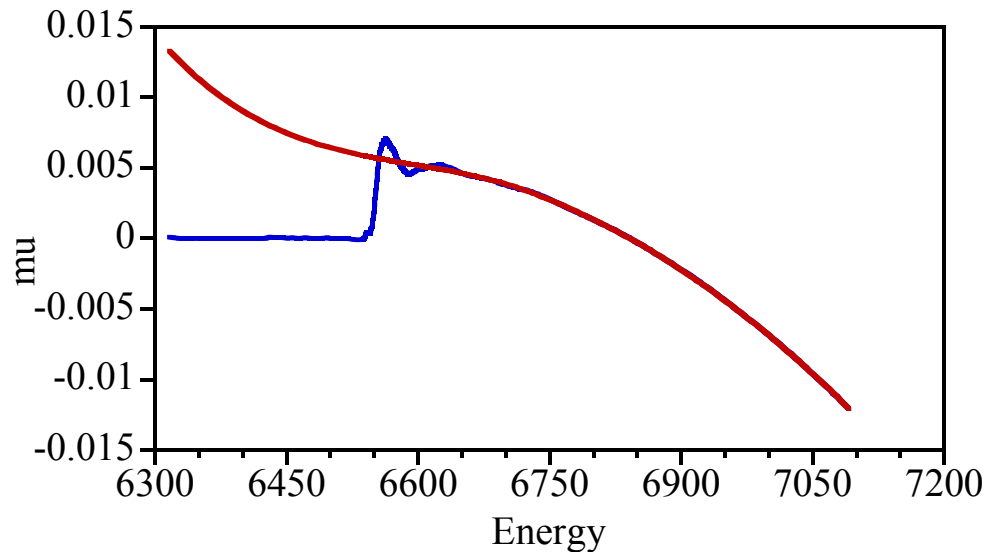
- Quantitative comparisons (e.g., titration) requires accurate normalization.
- Correction for various artifacts (self-absorption) requires accurate normalization.
- Common normalization procedures were developed for extracting EXAFS and do not necessarily work well for XANES.

Conventional normalization

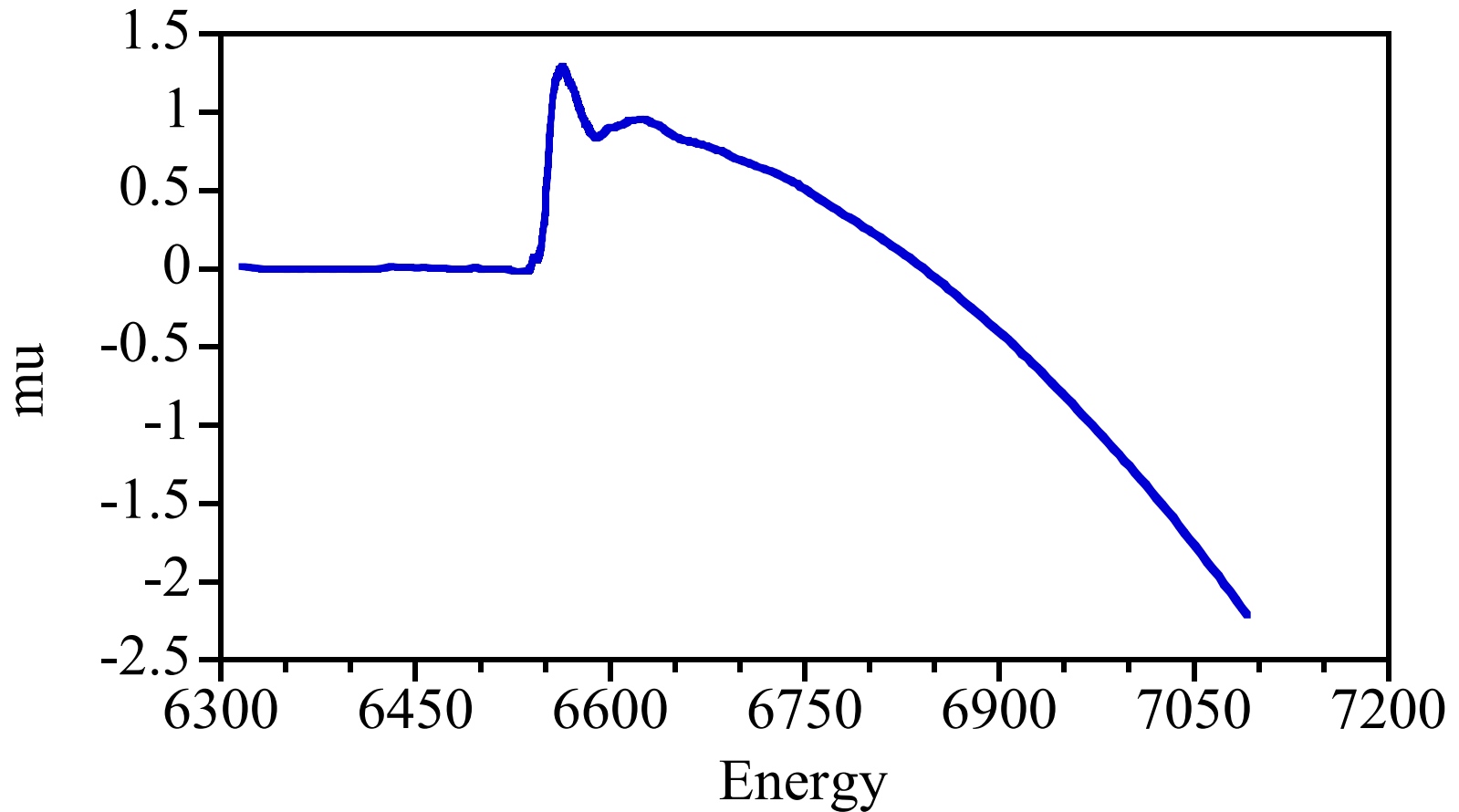


Pre-edge
subtraction
followed by
extrapolation

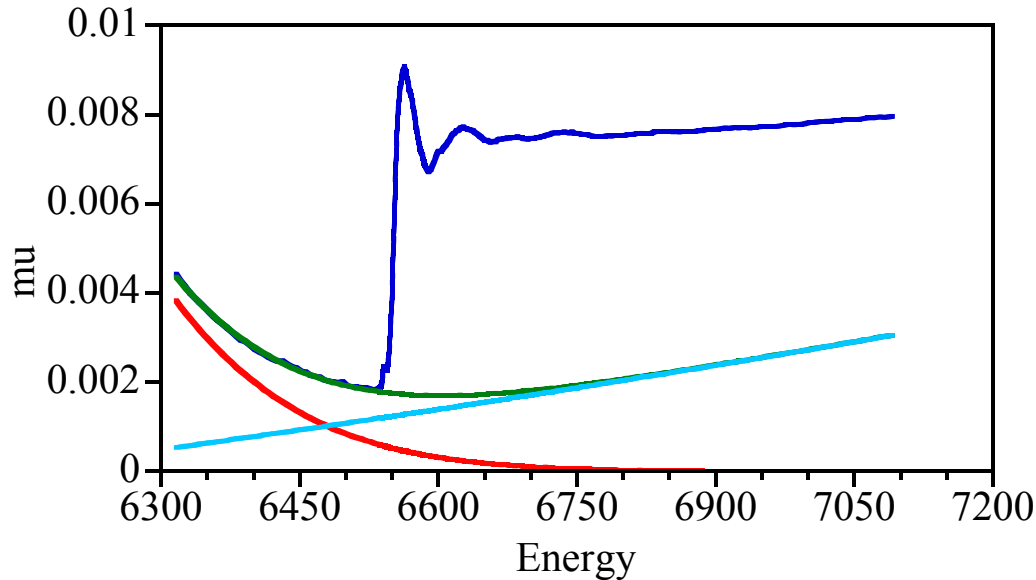
Post-edge (spline)
subtraction
followed by
extrapolation



Conventionally normalized data

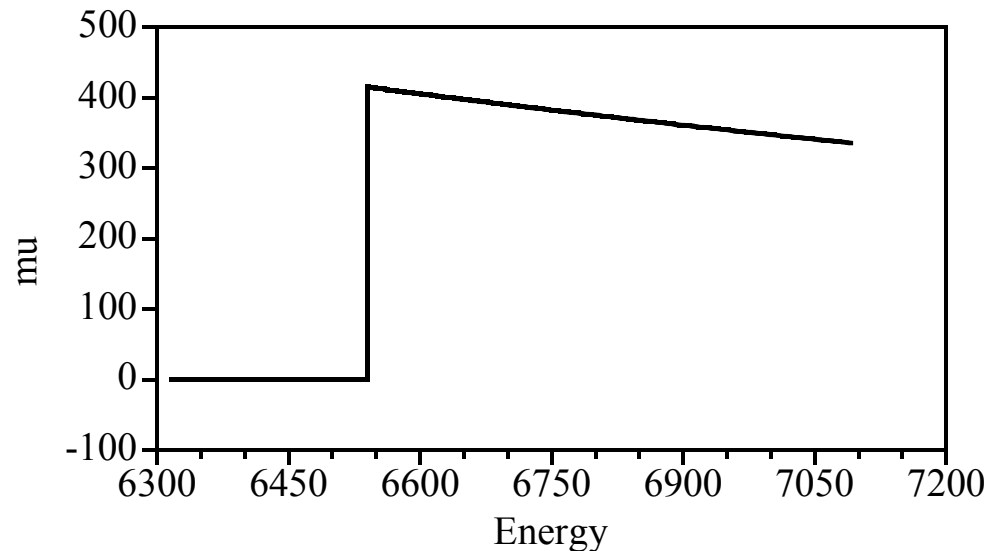


Alternative is to use a single background and tabulated cross-sections

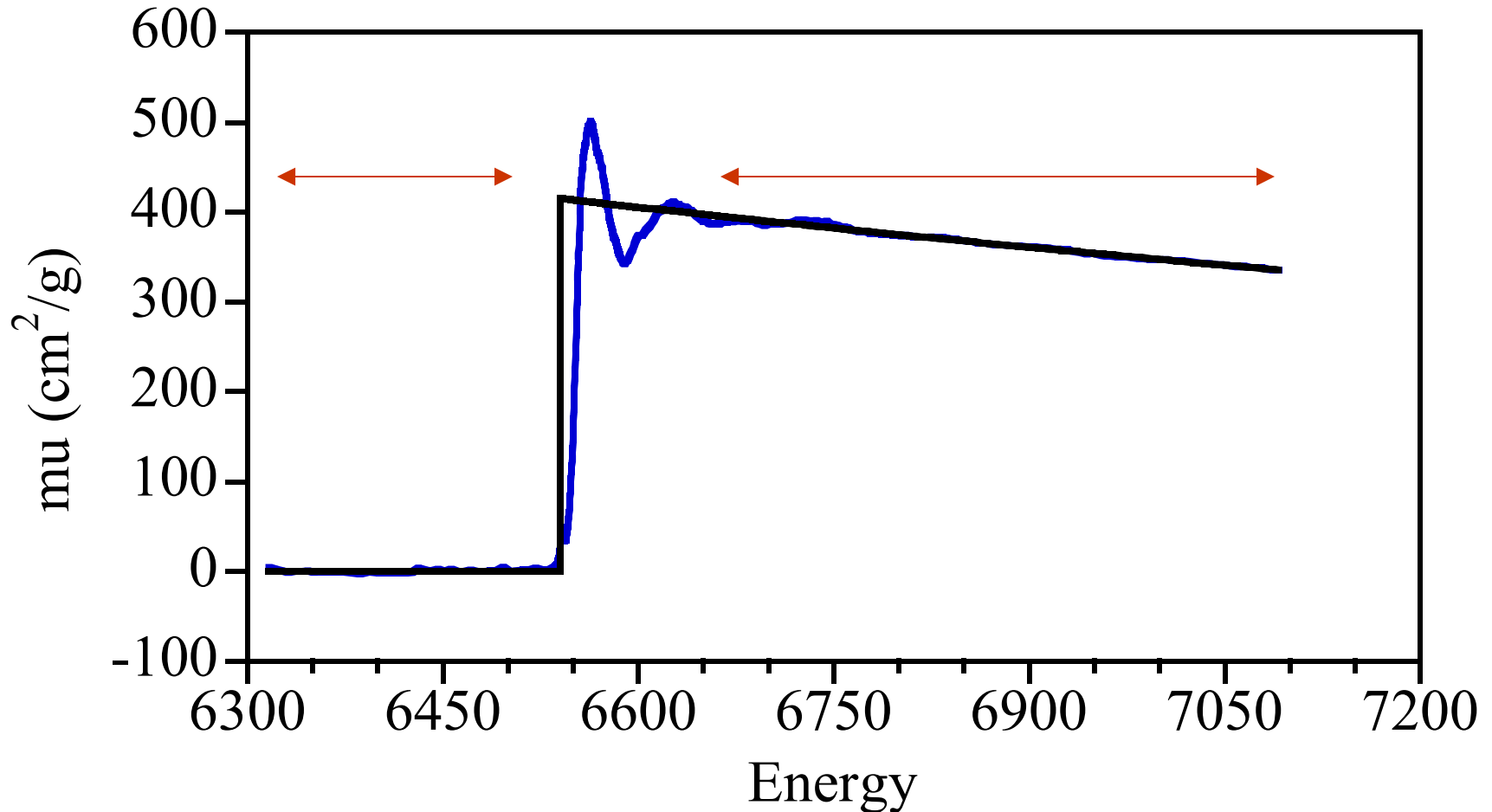


Polynomial +
complementary
error function
background fit to
all data

Fit to tabulated
McMaster values

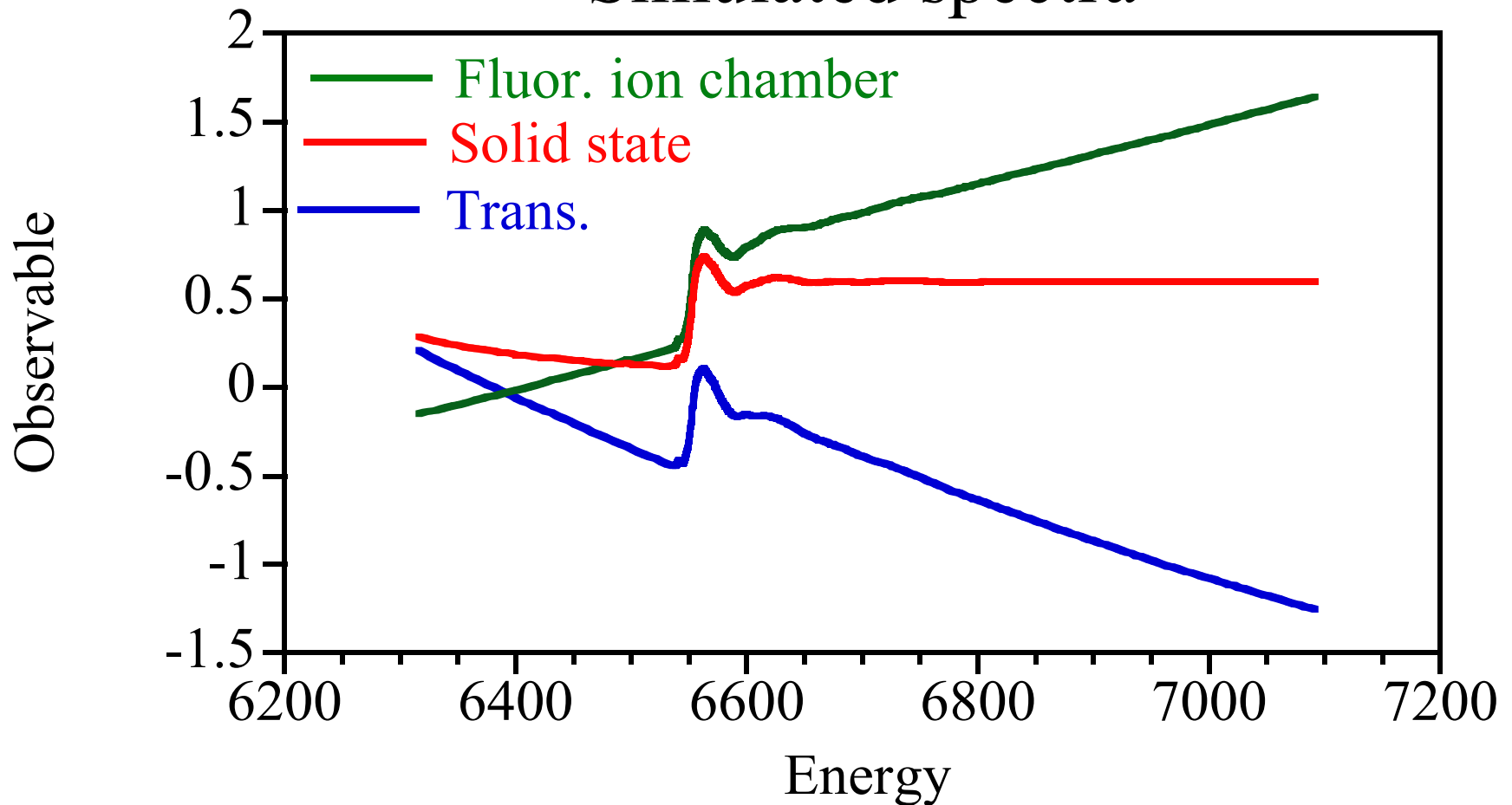


To avoid sensitivity to XANES features,
only fit data below and above edge

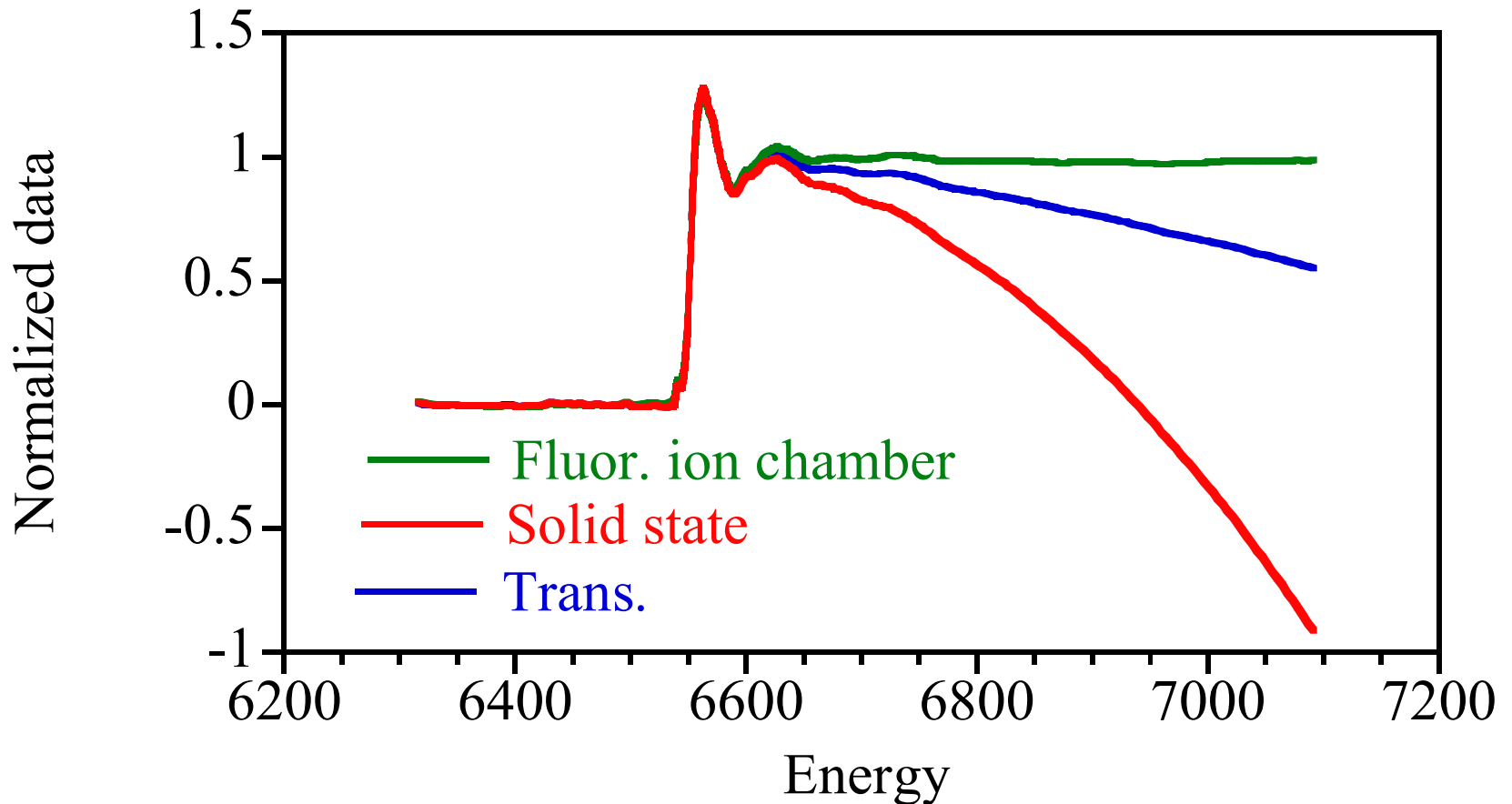


Ability to recover data using different backgrounds

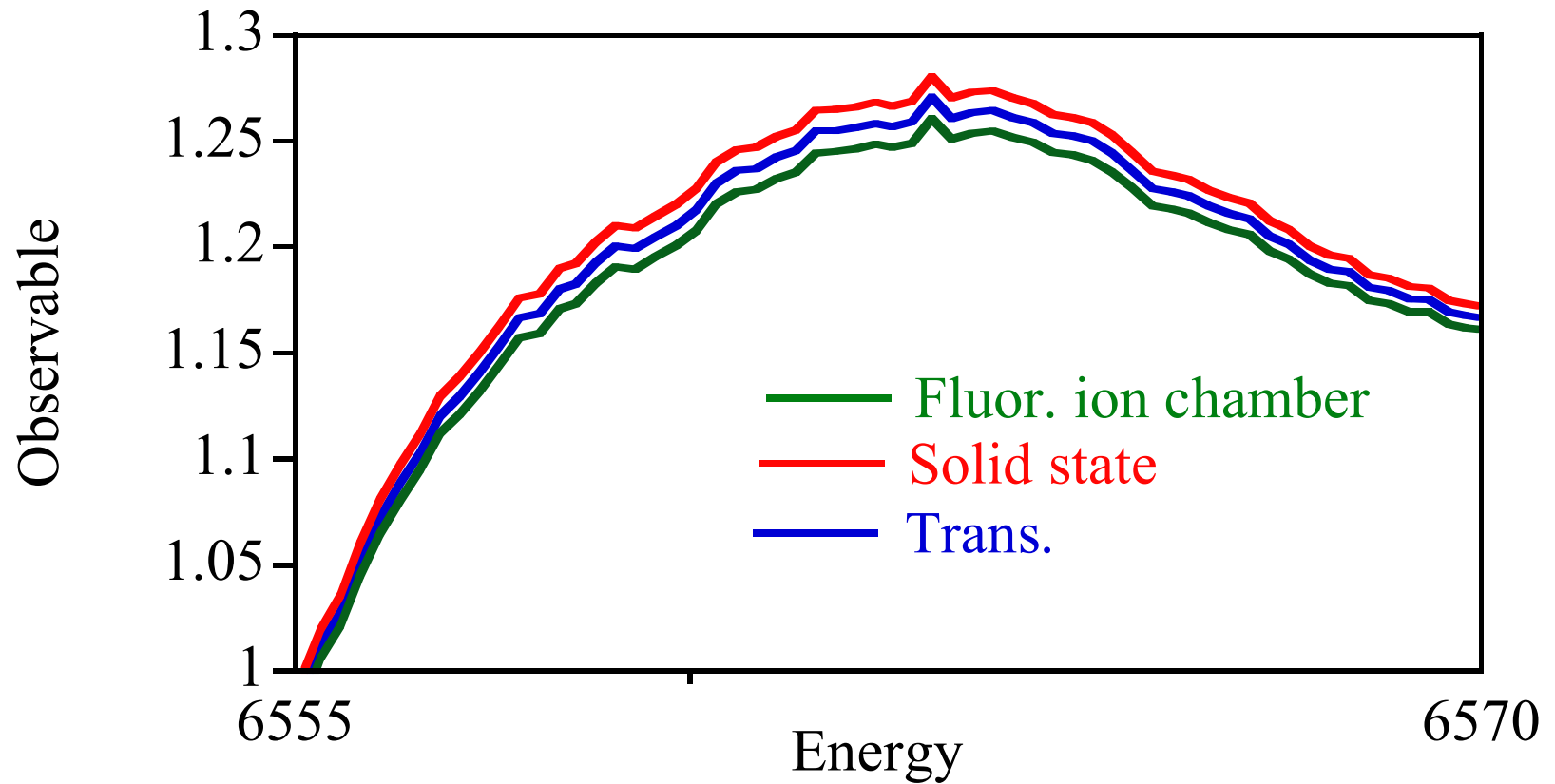
Simulated spectra



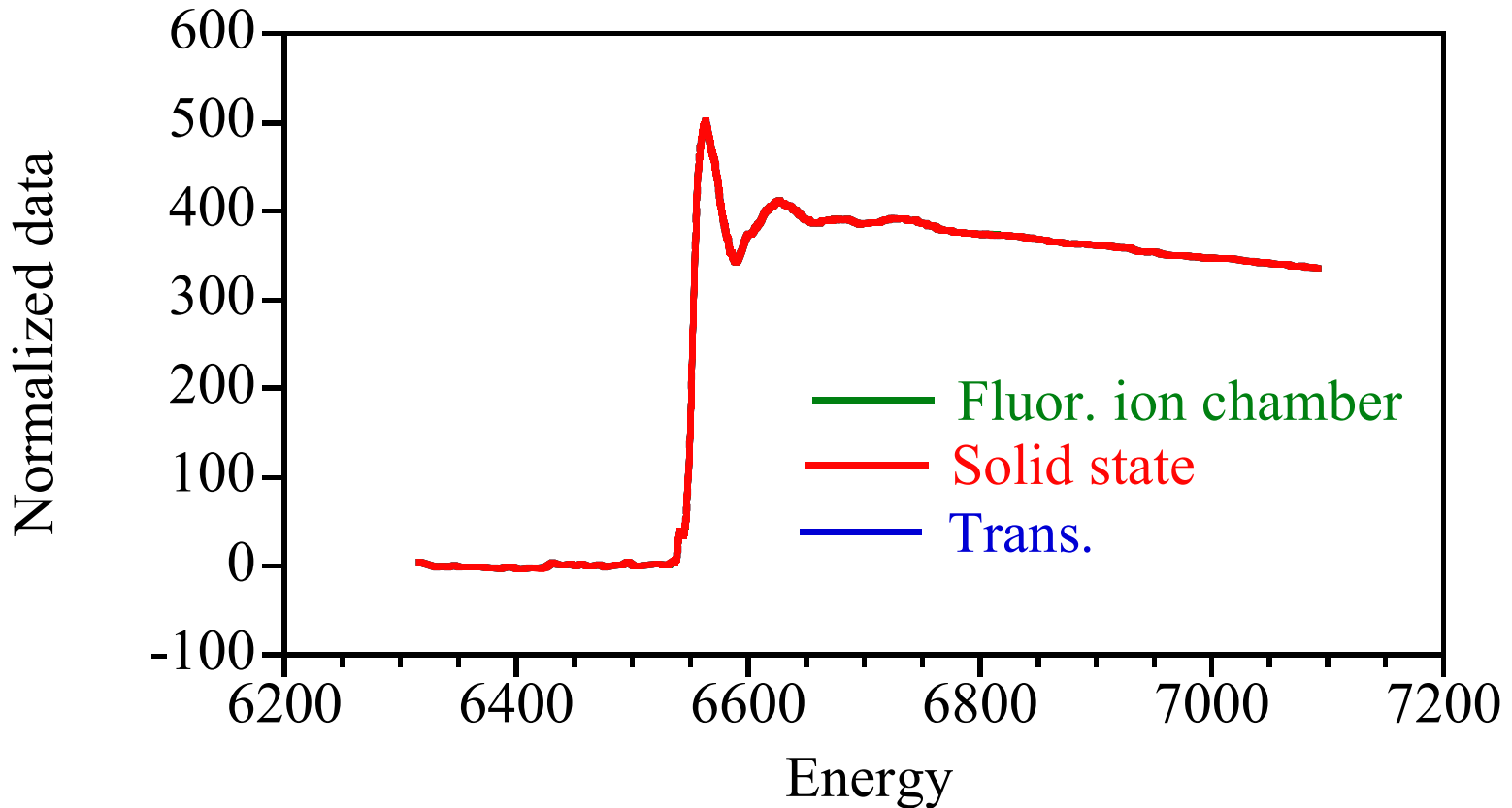
Conventional normalization is sensitive to background

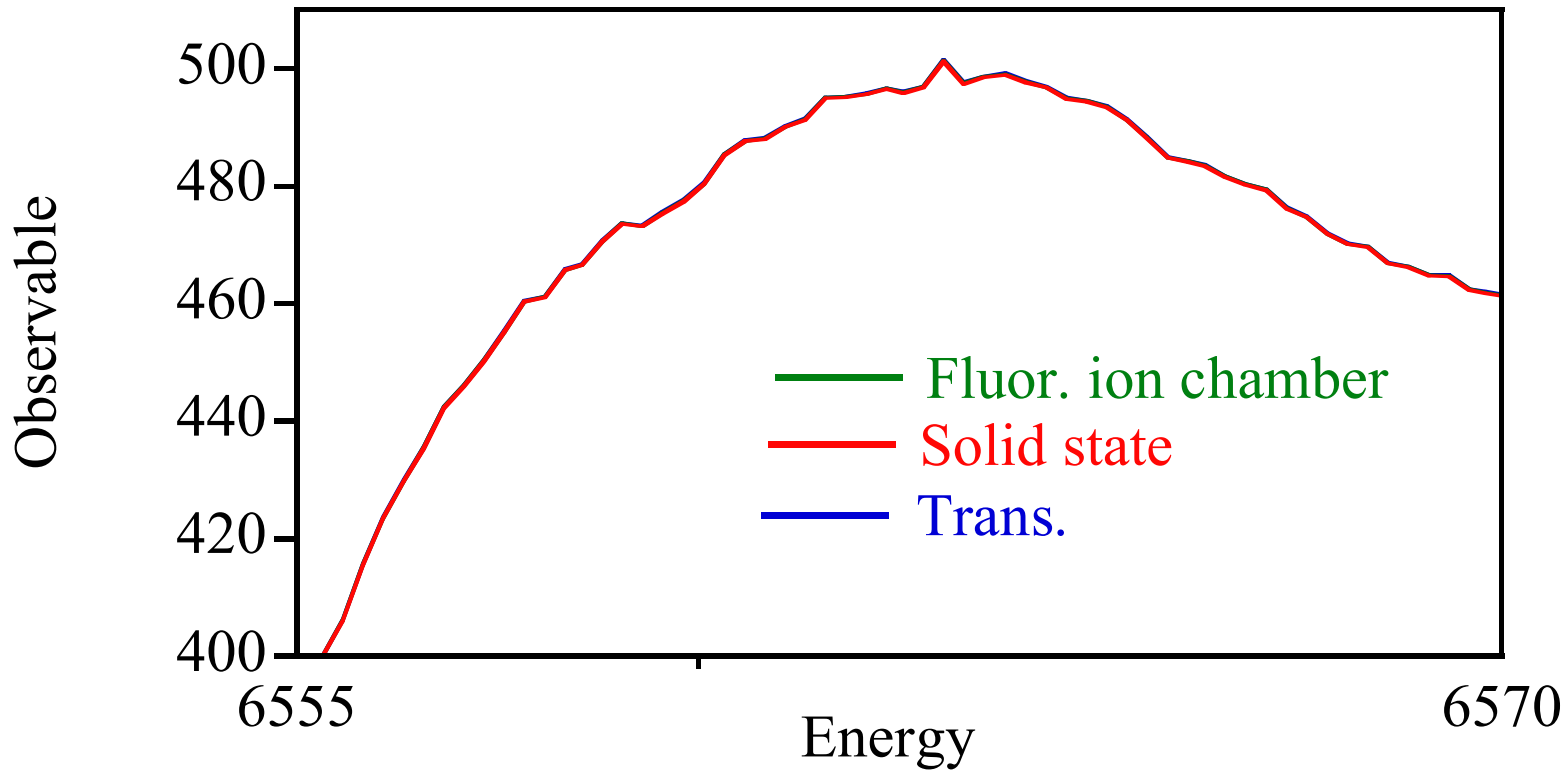


Errors in conventional normalization affect data even near edge

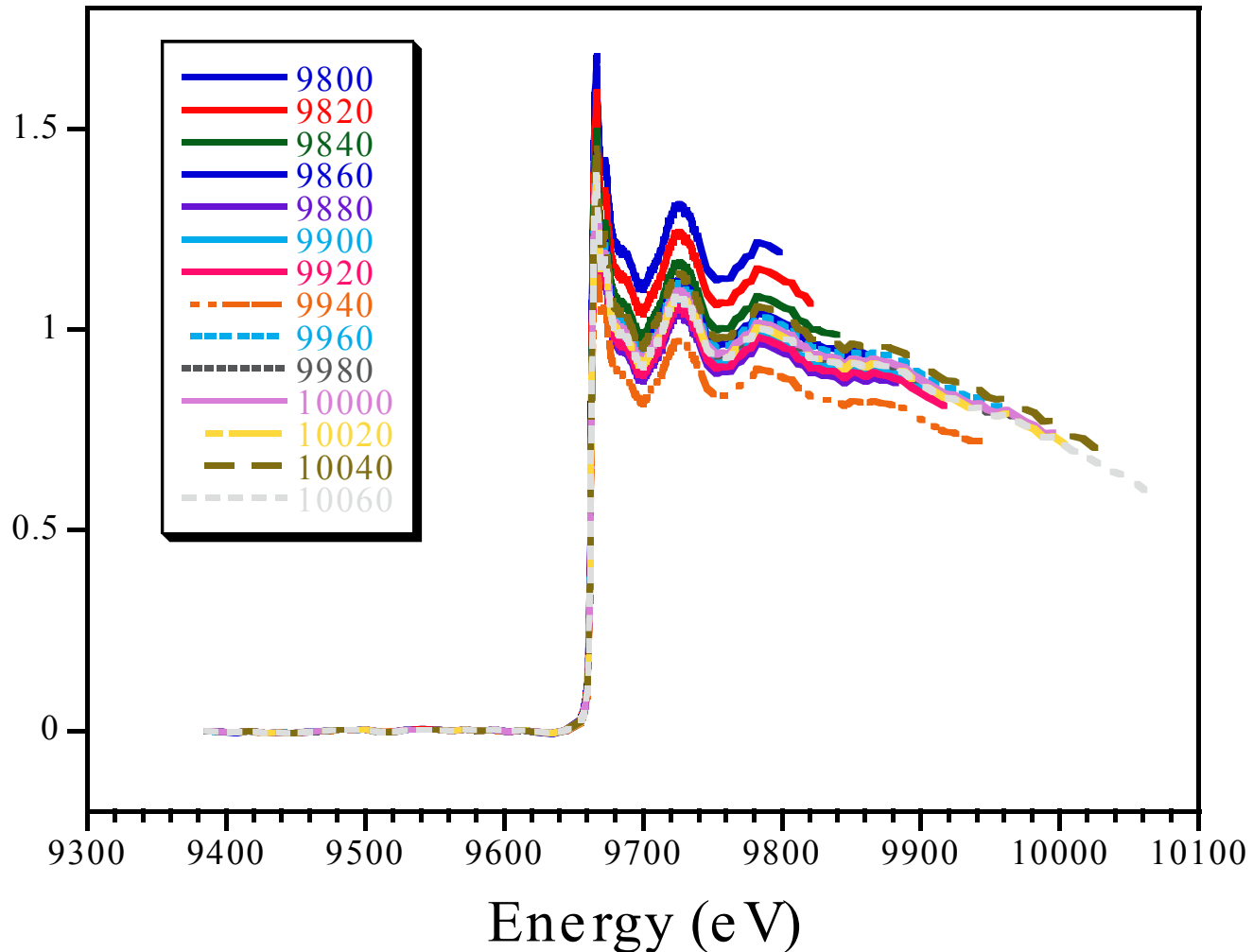


Proposed normalization is insensitive to background

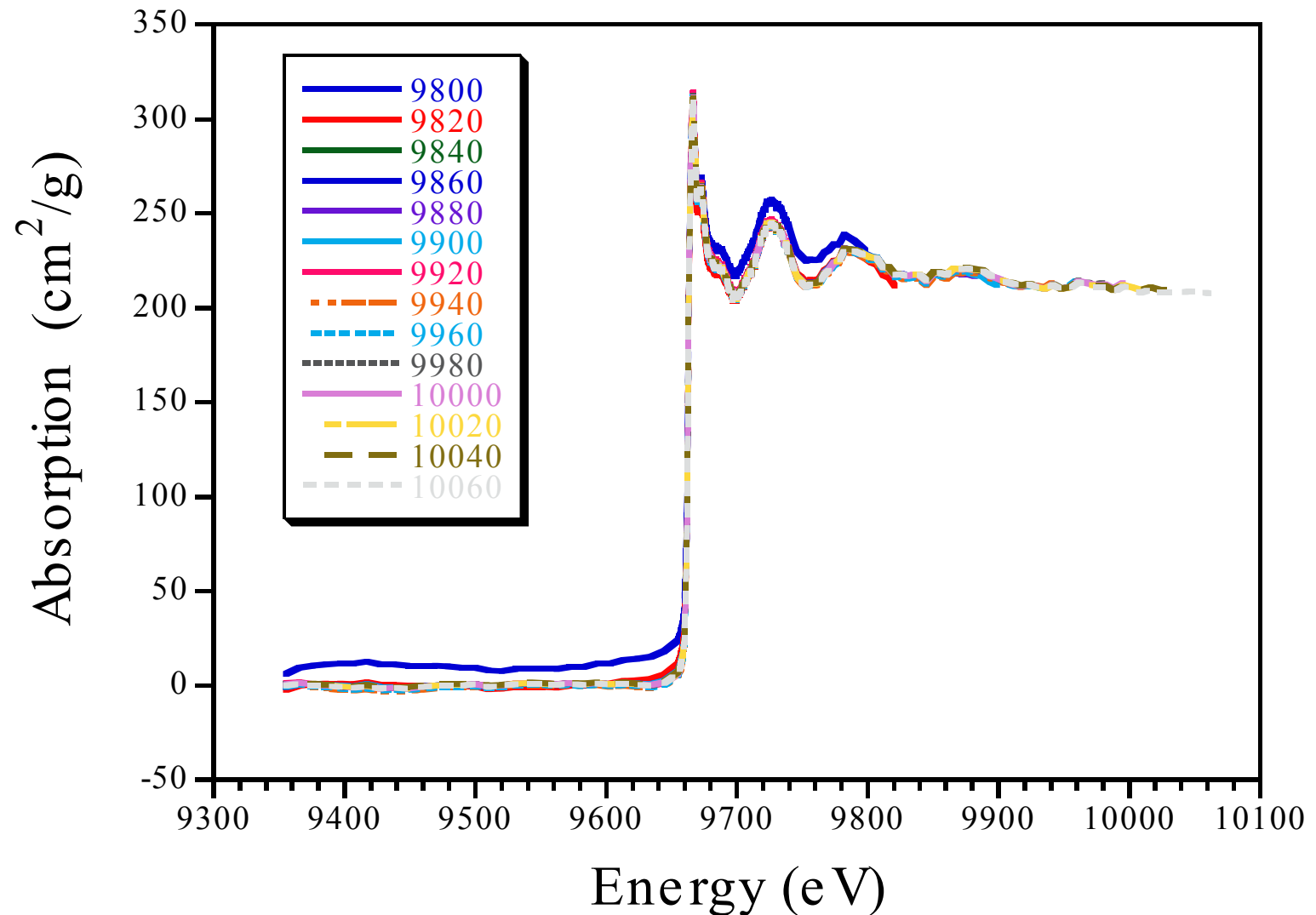




Conventional normalization is sensitive to range of data



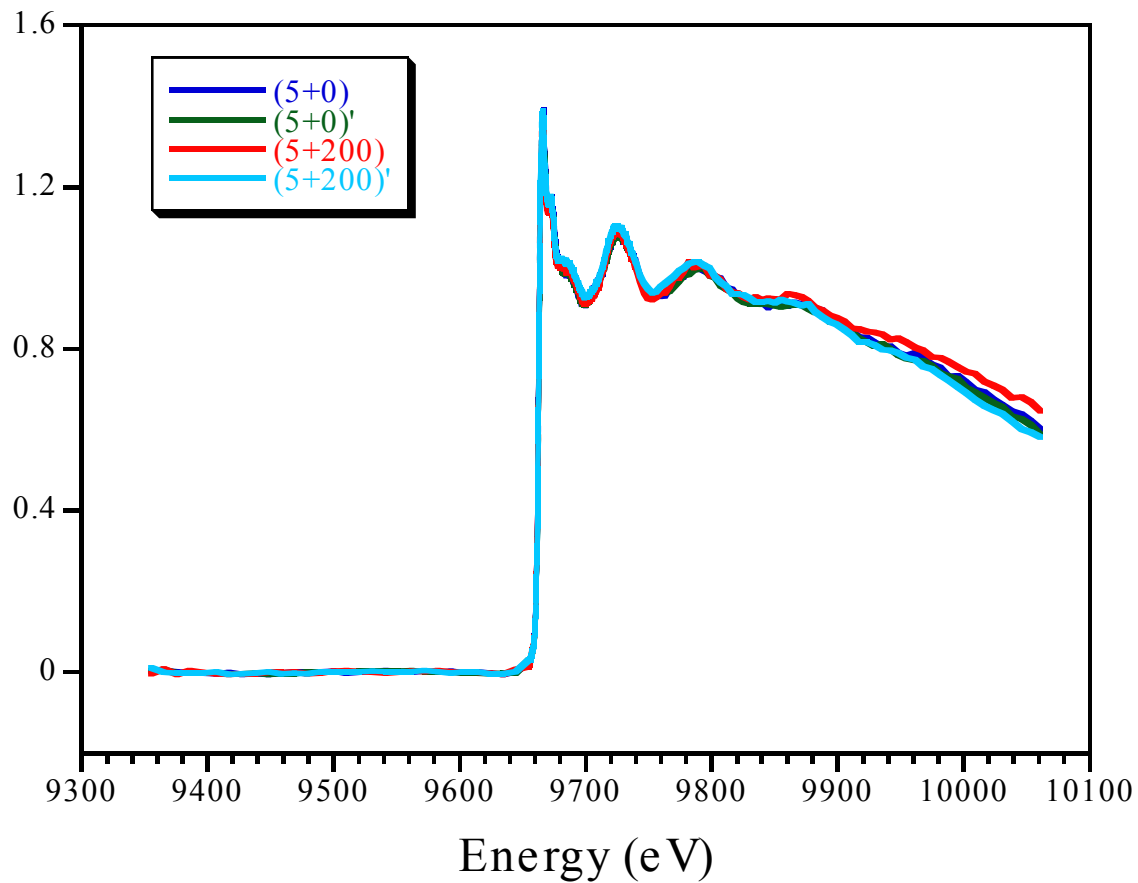
New method shows only slight sensitivity for $E_{\max} \geq \sim 150$ eV above edge



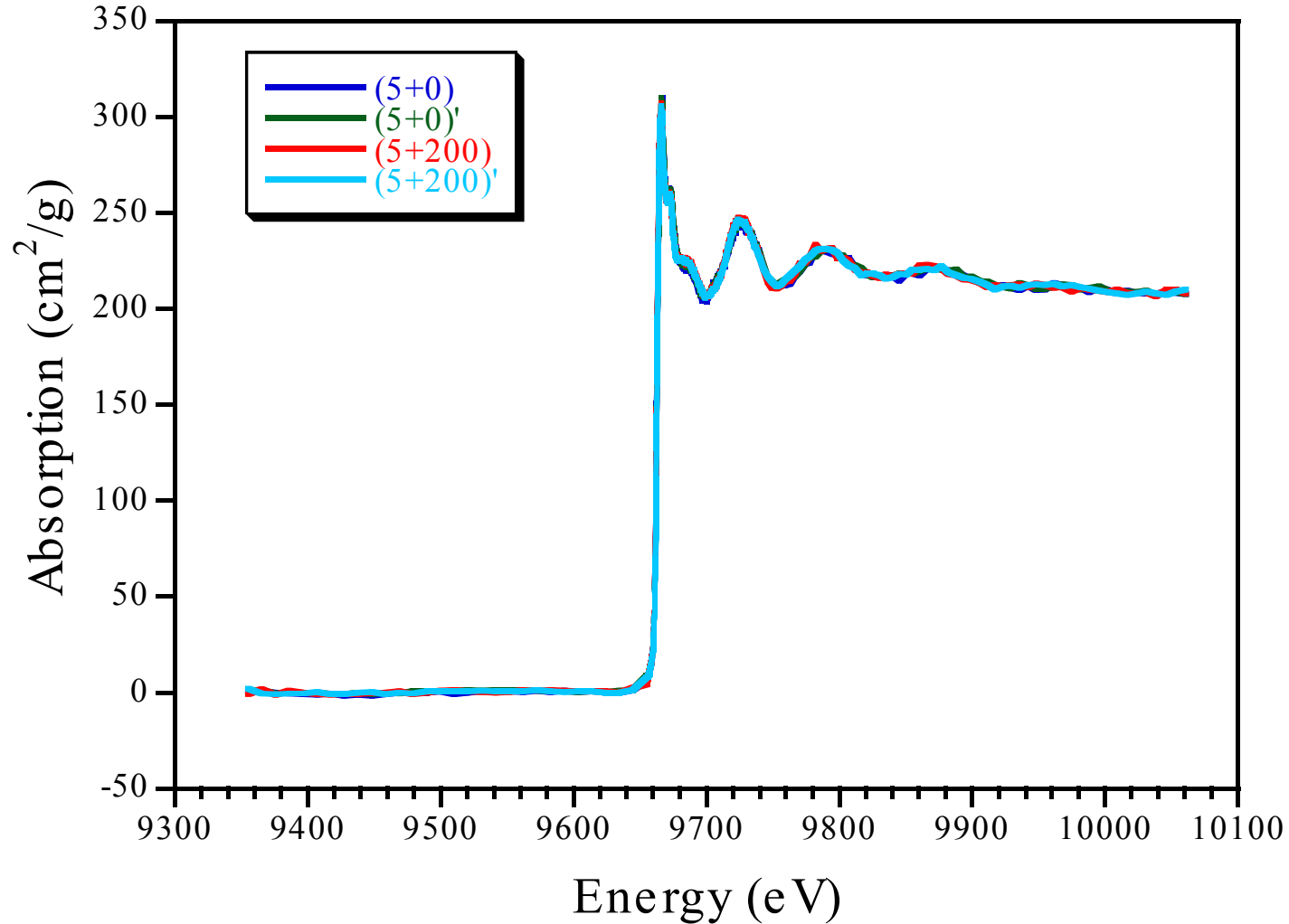
Although errors in conventional normalization are small, they affect conclusions

- $\text{Zn}(\text{SR})_4^{2-}$ dissociates in solution
- Complex can be forced to 100% $\text{Zn}(\text{SR})_4^{2-}$ by addition of excess RS^-
- Measure duplicate data for 5 mM $\text{Zn}(\text{SR})_4^{2-}$ with and without added RS^-

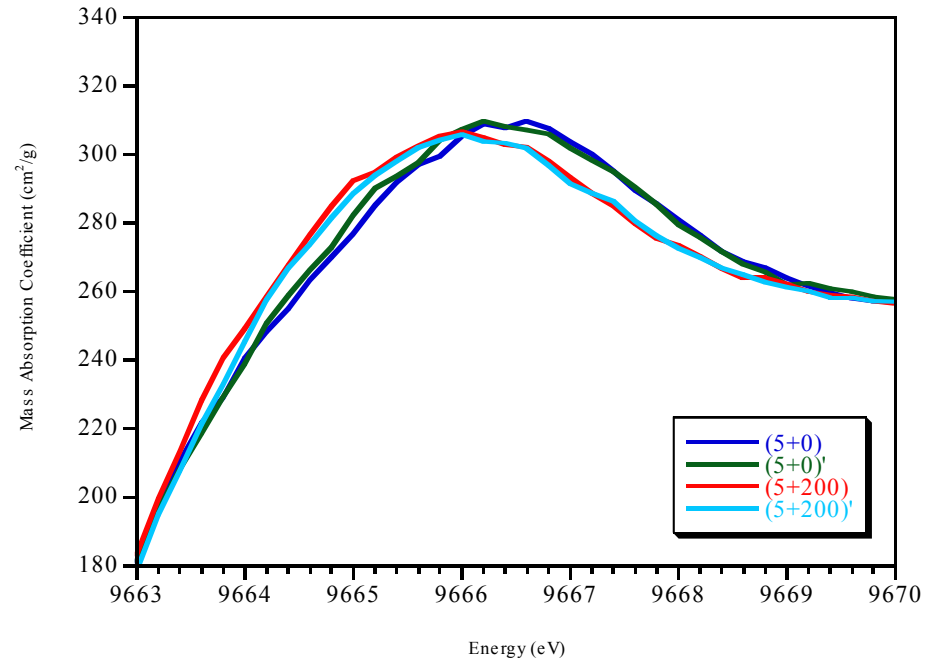
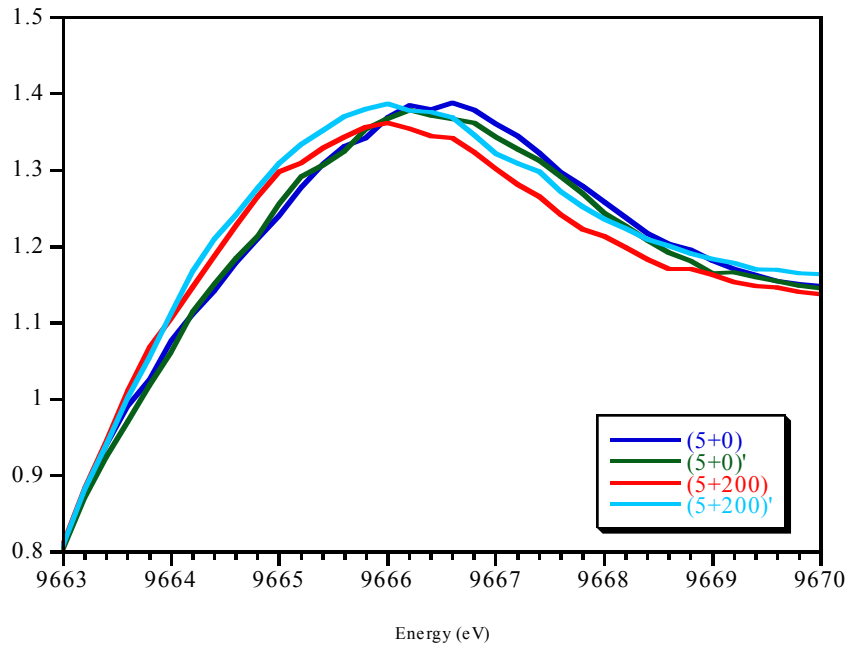
Conventional normalization



Proposed normalization method

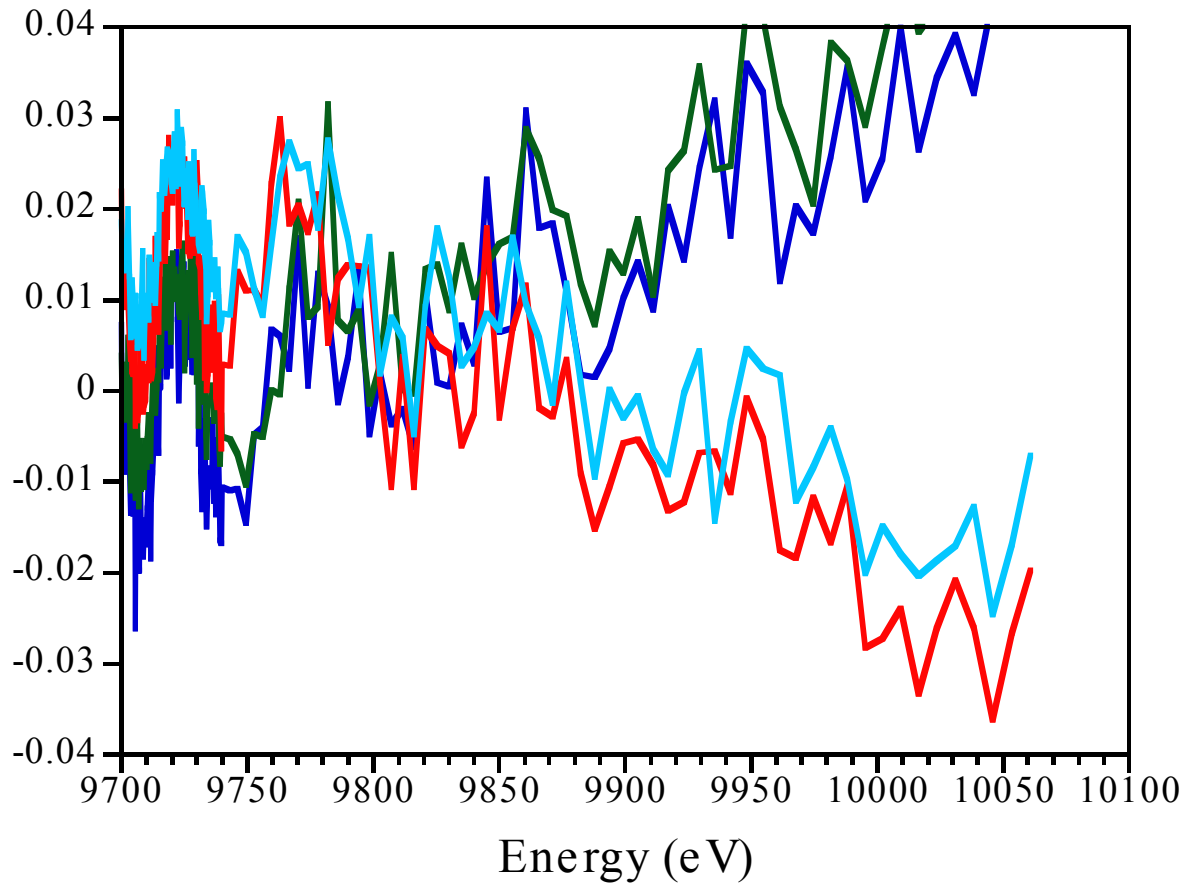


Variation in normalization obscures chemical effects

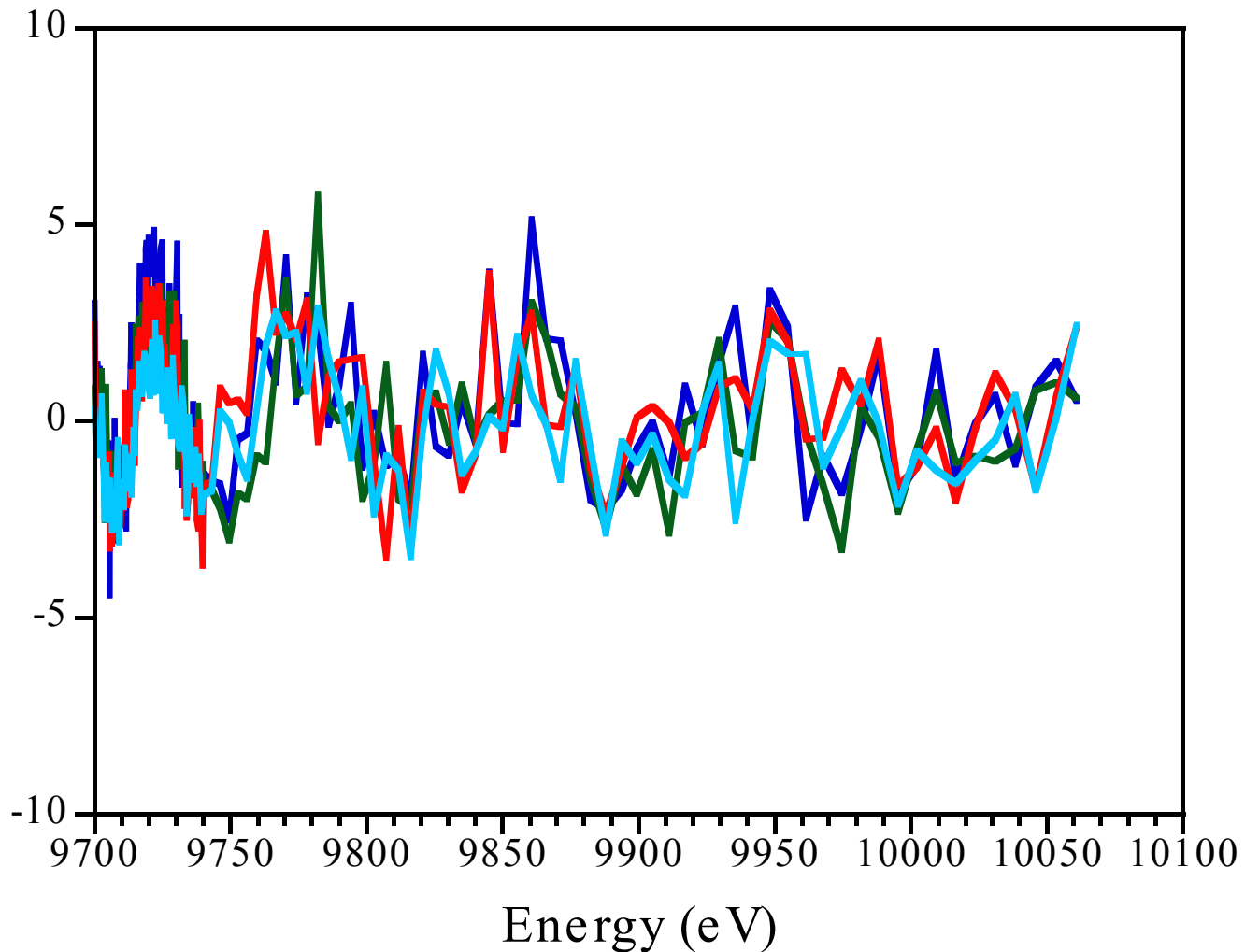


4 possible difference spectra – should all be the same

Conventional



With new normalization,
difference signal is detectable



Acknowledgements

- Kimber Clark
- David Tierney
- Geoff Waldo
- Tsu-Chien Weng
- NIH

International XAFS Society

Report on error analysis. See

<http://ixs.csrri.iit.edu/>

Public comment period – please review and, if desired, comment.